

Therapeutic Inhibition of ENaC With a Lung-Targeted RNAi Molecule Delivery Platform Preserves Normal Mucus Clearance in a Mucostatic Sheep Model of Cystic Fibrosis

Anthony Nicholas¹, Juan R. Sabater², Tao Pei¹, Xiaokai Li¹, Agnieszka Glebocka¹, Holly Hamilton¹, Julia Hegge¹, Zach Trilling¹, Thomas Schluep³, Matthias A. Salathe⁴, Burton F. Dickey⁵, Steven M. Rowe⁶, Marcus A. Mall^{7,8}, Erik W. Bush¹

¹ Arrowhead Pharmaceuticals Inc., Madison, WI, United States; ² Mount Sinai Medical Center, Miami Beach, FL, United States; ³ Arrowhead Pharmaceuticals Inc., Pasadena, CA, United States; ⁴ University of Kansas, Kansas City, KS, United States; ⁵ Department of Pulmonary Medicine, University of Texas M. D. Anderson Cancer Center, Houston, TX, United States; ⁶ University of Alabama at Birmingham, Birmingham, AL, United States; ⁷ Department of Pediatric Pulmonology, Immunology and Intensive Care Medicine, Charité-Universitätsmedizin Berlin, Berlin, Germany; ⁸ Berlin Institute of Health (BIH), Berlin, Germany

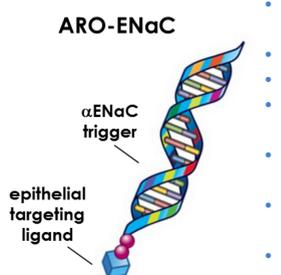
RATIONALE

In cystic fibrosis (CF), increased epithelial sodium channel (ENaC) activity accompanies loss of CFTR channel function and contributes to airway dehydration and mucoobstruction¹. Loss-of-function ENaC alleles increase mucociliary clearance (MCC) and modify CF lung disease to milder phenotypes³, while gain-of-function alleles contribute to the pathogenesis of atypical CF⁴. Despite strong genetic and functional⁵ validation of the target, clinical development of inhaled small molecule ENaC inhibitors has been limited by their short duration of action and renal side effects⁶. Using Arrowhead's Targeted RNAi Molecule (TRiM™) technology, we developed ARO-ENaC (an optimized RNAi trigger against α ENaC mRNA paired with a pulmonary epithelial targeting ligand), which has previously been shown to durably and selectively silence α ENaC expression in the rodent lung⁷. In the current study, we evaluated the effects of α ENaC silencing on large animal lung physiology. Mucociliary clearance was measured in ARO-ENaC-treated normal sheep and in sheep challenged with neutrophil elastase, an ENaC-activating protease that promotes airway surface liquid depletion, increased mucus secretion and mucostasis mimicking CF lung disease.

METHODS

- Male Sprague Dawley rats received aerosolized ARO-ENaC conjugate or α ENaC trigger alone, delivered via jet nebulizer and nose-only inhalation. One week after dosing, total RNA was isolated from whole lung homogenates and mRNA expression analyzed by qPCR.
- Baseline mucociliary clearance (MCC) in conscious, nasally intubated female sheep (n=3 per group) was measured by inhalation of aerosolized technetium-labeled sulfur colloid (^{99m}Tc-SC) followed by gamma imaging at five minute intervals for two hours.
- Aerosolized ARO-ENaC conjugate or α ENaC trigger alone was delivered to nasally intubated sheep (either one dose or three daily doses). Follow-up mucociliary clearance scans were performed weekly at 7, 14, 21 and 28 days post-dose.
- For the impaired MCC model, human neutrophil elastase (HNE) was administered by inhalation immediately prior to gamma imaging⁸.
- Deposited pulmonary doses were calculated using the respiratory minute volume, aerosol concentration, exposure time, body weight and deposition fraction (10% for rat; 25% for sheep).

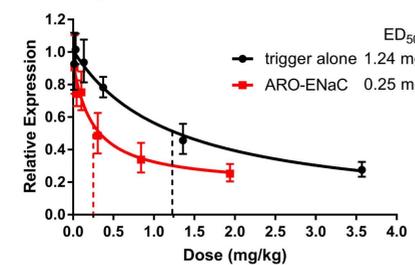
TRiM™ platform

- 
- Rules and algorithms allow selection of optimized RNAi trigger sequences
 - Limit cross-reactivity with off-target genes
 - Maximize innate stability
 - Rational use and placement of modifying chemistries
 - Active endosomal escape chemistries not required
 - Targeting ligands and linker chemistries improve delivery to target tissues
 - Integrin α v β 6 targeting moiety facilitates pulmonary epithelial uptake and endocytosis of triggers

RESULTS

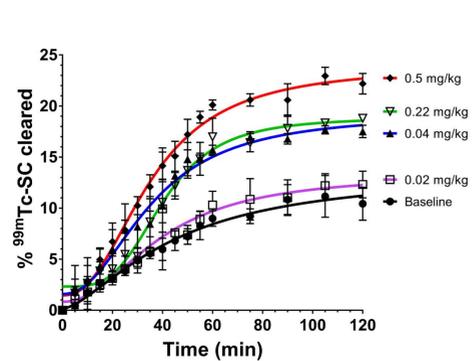
Epithelial targeting ligand increases potency of whole lung α ENaC mRNA silencing following inhaled aerosol dosing in rats

Rat whole lung α ENaC expression
Day 1 inhaled ARO-ENaC; Day 8 sacrifice

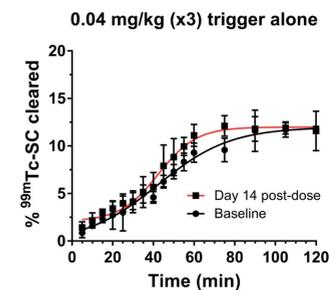
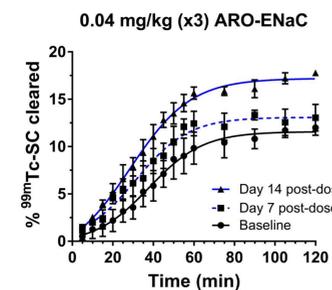


- Rats received a single inhaled aerosol dose of ARO-ENaC conjugate (RNAi trigger + ligand) or trigger alone. Whole lung α ENaC mRNA expression was evaluated one week post-dose
- ARO-ENaC's epithelial targeting ligand increases potency of trigger

Dose-dependent acceleration of mucociliary clearance (MCC) in normal sheep two weeks after ARO-ENaC inhalation

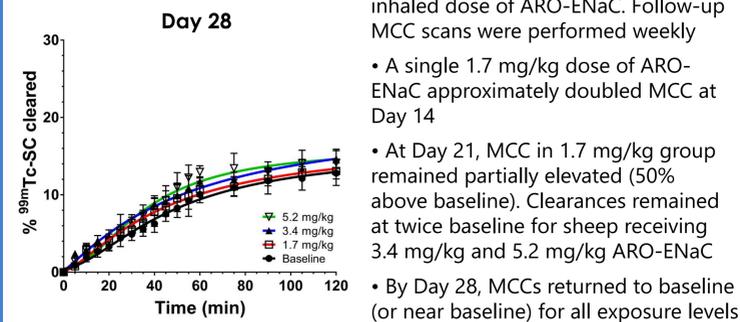
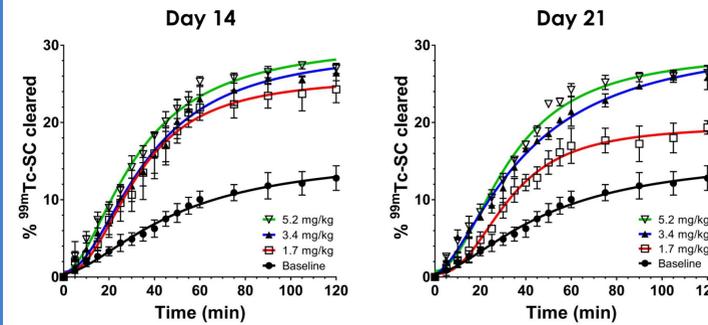


- At baseline, sheep cleared approximately 10-12% of inhaled tracer
- On Days 1-3, ARO-ENaC was inhaled at 0.5, 0.22, 0.04, and 0.02 mg/kg deposited dose levels
- On Day 17, accelerated MCC was observed at all doses: +113% above baseline (0.5 mg/kg), +80% (0.22 mg/kg), +68% (0.04 mg/kg) and +18% (0.02 mg/kg)



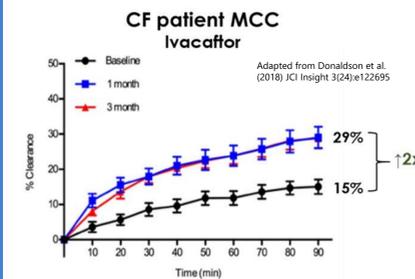
- Accelerated MCC could be observed one week after ARO-ENaC inhalation
- Inhaled RNAi trigger without targeting ligand did not accelerate MCC

A single inhaled dose of ARO-ENaC accelerates mucociliary clearance in normal sheep up to three weeks

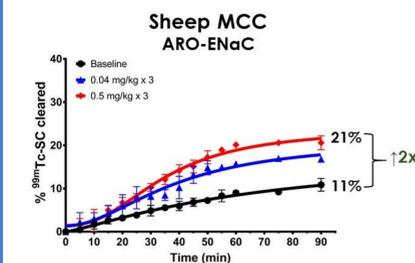


- On Day 1, sheep received a single inhaled dose of ARO-ENaC. Follow-up MCC scans were performed weekly
- A single 1.7 mg/kg dose of ARO-ENaC approximately doubled MCC at Day 14
- At Day 21, MCC in 1.7 mg/kg group remained partially elevated (50% above baseline). Clearances remained at twice baseline for sheep receiving 3.4 mg/kg and 5.2 mg/kg ARO-ENaC
- By Day 28, MCCs returned to baseline (or near baseline) for all exposure levels

α ENaC silencing improves mucociliary clearance with a benefit similar in magnitude to ivacaftor in G551D-CFTR CF patients

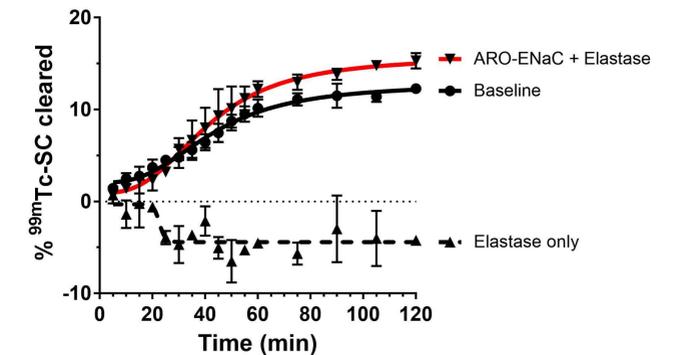


- As previously shown by Donaldson et al.⁹, lung MCC doubled in G551D-CFTR CF patients within one month of initiation of ivacaftor treatment (150 mg, BID)
- In ivacaftor-treated patients, accelerated MCC correlated with improved lung function, measured by FEV₁



- Similarly, three daily inhaled deposited doses of 0.5 mg/kg ARO-ENaC doubled lung MCC in normal sheep two weeks after the third dose

ARO-ENaC preserves airway physiology in a sheep disease model of impaired mucociliary clearance



- Neutrophil elastase cleaves and activates near-silent pulmonary epithelial ENaC channels¹⁰ and may also increase mucus secretion, contributing to airway surface liquid depletion, mucus hyperconcentration and impaired clearance
- When administered to sheep via inhalation, neutrophil elastase caused complete mucostasis (dotted line), with no tracer cleared over two hours
- Sheep receiving three inhaled doses of 0.5 mg/kg ARO-ENaC two weeks prior to neutrophil elastase challenge (red line) were fully protected from mucostasis, maintaining lung clearance approximately 25% above their pretreatment baselines

CONCLUSIONS

- Arrowhead's pulmonary epithelial TRiM™ delivery platform increases potency of RNAi trigger-mediated silencing of α ENaC mRNA, durably reducing whole lung gene expression in rodents
- In normal sheep, inhaled ARO-ENaC produces dose-dependent acceleration of mucociliary clearance, a lung physiology endpoint linked to pulmonary function in CF patients
- Inhaled ARO-ENaC preserves lung clearance in a sheep disease model of mucostasis caused by challenge with the ENaC-activating protease neutrophil elastase
- ARO-ENaC offers a new renal-sparing, genotype-agnostic mucokinetic therapy for all CF patients, with an extended duration of action that should minimize treatment burden. IND/CTA-enabling studies are in process to support regulatory filings for first-in-human studies
- Arrowhead is expanding the pulmonary delivery platform to address additional disease targets, particularly those that are inaccessible to traditional small molecule or antibody approaches

REFERENCES

- Hobbs C et al. (2013) J Physiol 591:4377-4387
- Kerem E et al. (1999) N Engl J Med 341:156-161
- Agrawal P et al. (2017) Am J Respir Cell Mol Biol 57:711-720
- Rauh R et al. (2010) J Physiol 588: 1211-1225
- Mall M et al. (2004) Nat Med 10(5):487-493
- O'Riordan T et al. (2014) J Aerosol Med Pulm Drug Deliv 27:200-208
- Bush E et al. (2018) NACFC
- Sabater J R et al. (2009) Am J Respir Crit Care Med 179:A1440
- Donaldson et al. (2018) JCI Insight 3(24):e122695
- Caldwell et al. (2005) Am J Physiol Lung Cell Mol Physiol 288:L813-L819